

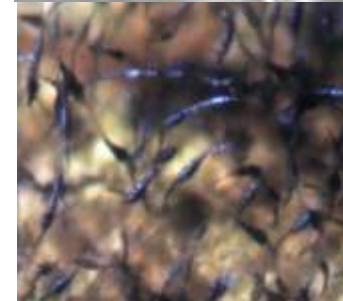
Population genetics

—

Consequences on early blight disease

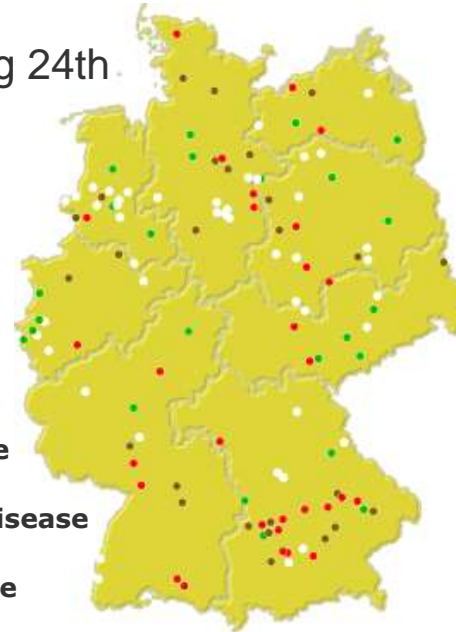
J. Leiminger, G. Bahnweg, H. Hausladen

*Euroblight Workshop
4.-7. May 2010*





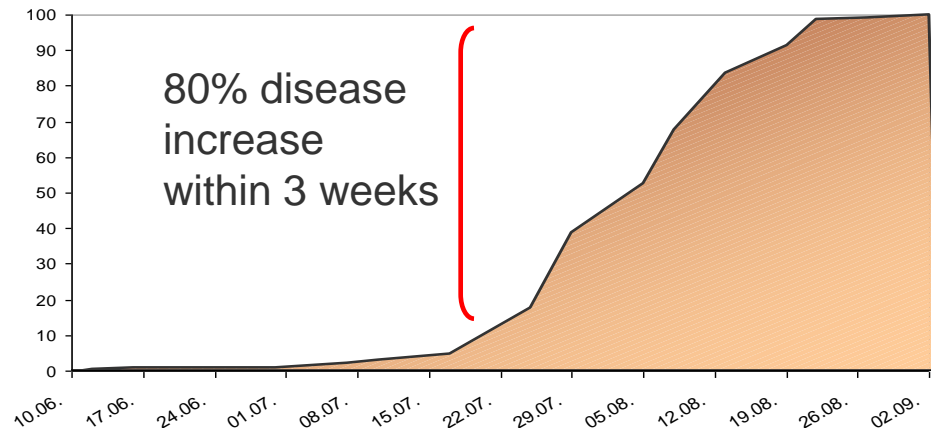
2009, Aug 24th



- No disease
- Low disease
- Moderate disease
- High disease



necrotic leaf area (%)



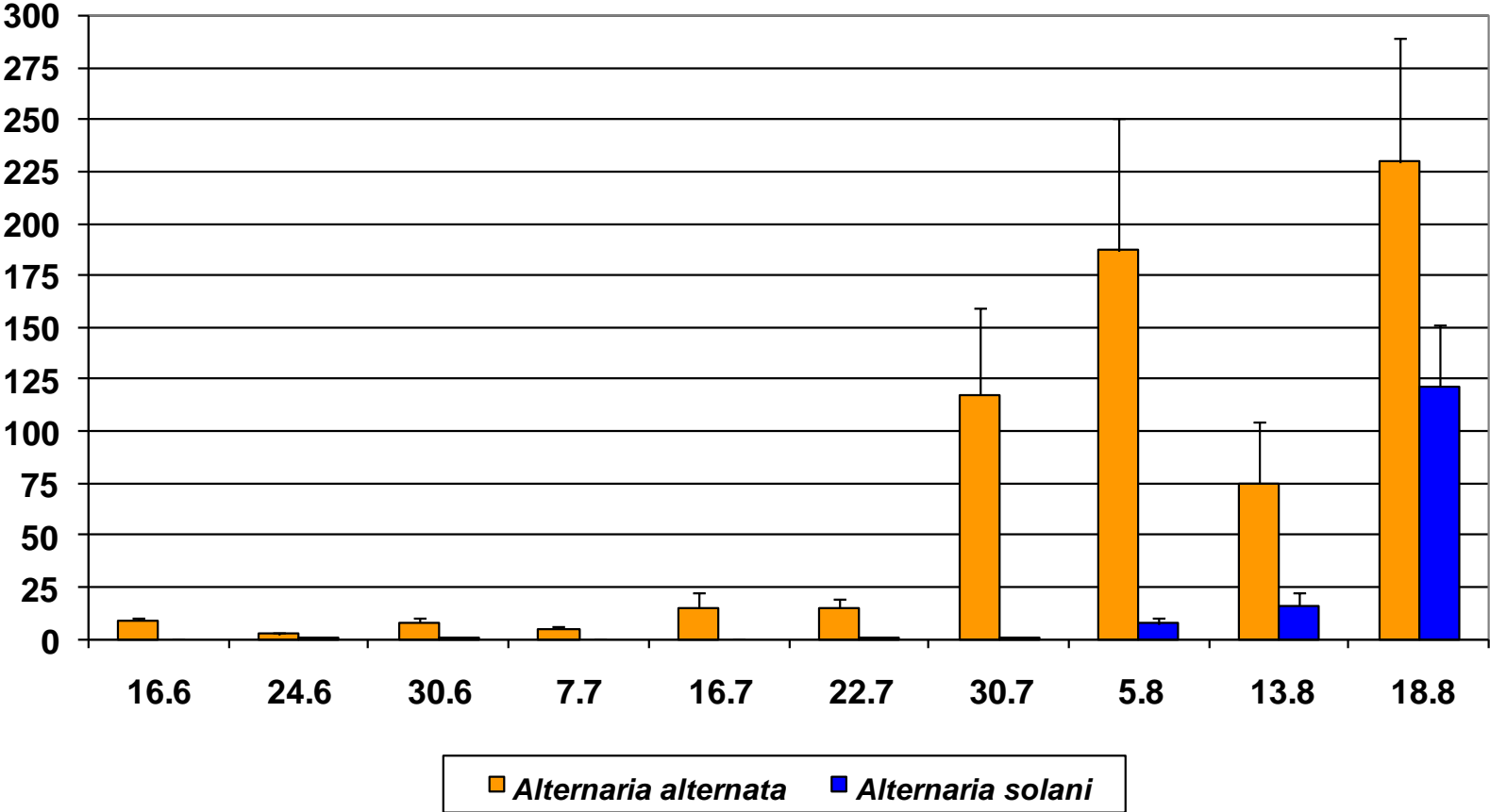
80% disease increase within 3 weeks



sampling	location	A.a	A.s.
09. Jun	Gundhöring	✓	✓
14. Jun	Laberweinting	/	✓
13. Jun	Aiterhofen	✓	/
17. Jun	Worms	✓	/
12. Jul	Cappel-Cloppenburg	✓	/
14. Jul	Dessau	✓	✓
19. Jul	Salching	✓	✓
21. Aug	Meppen	✓	✓
21. Aug	Meppen	✓	✓
21. Aug	Rupenest	✓	✓
21. Aug	Niewe-Schanz	✓	✓
25. Aug	Lichtenau	✓	✓
05. Sep	Pönning	✓	✓
05. Sep	Landshut	✓	✓
06. Sep	Aiterhofen	✓	✓
09. Sep	Pulling	✓	✓

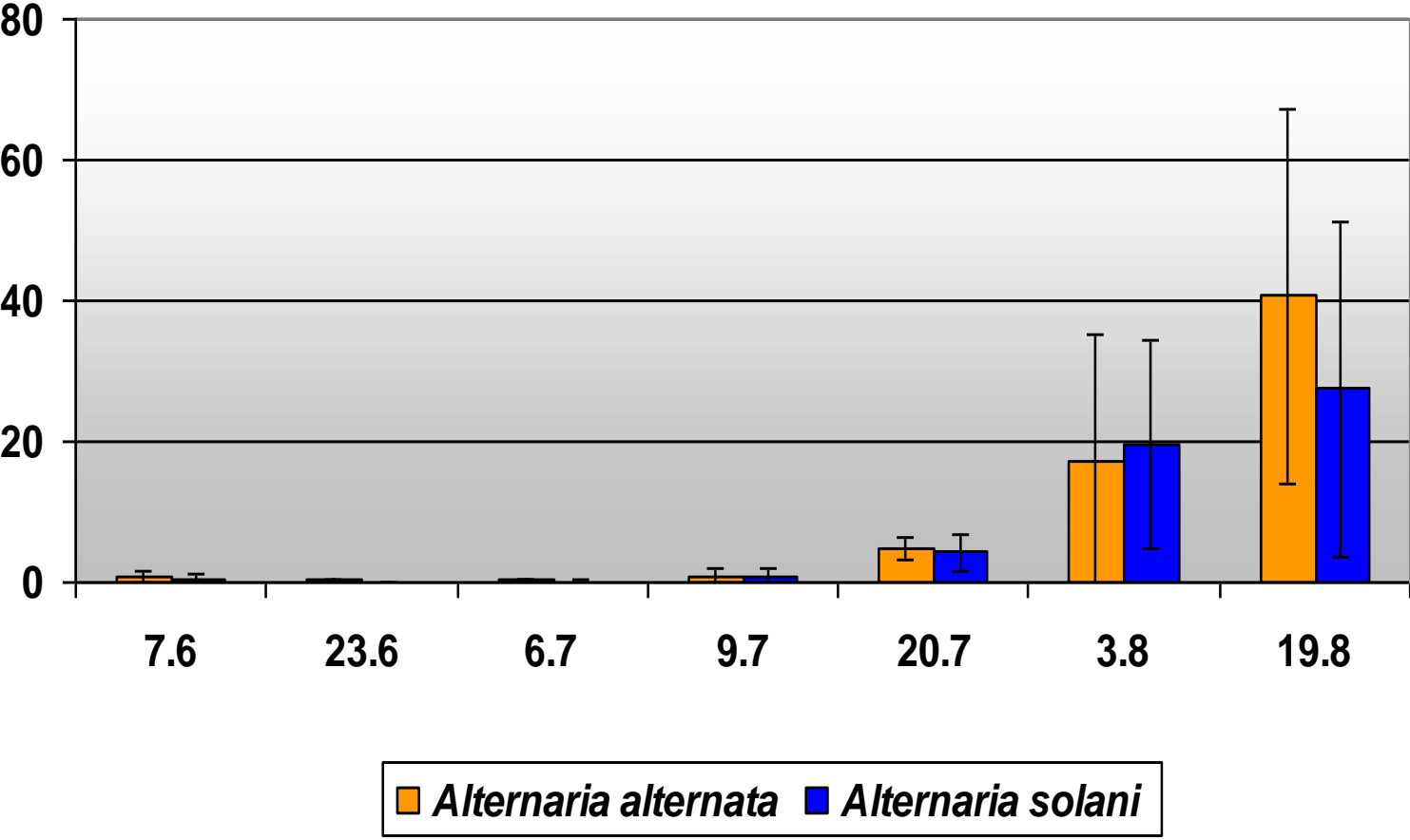
2003, Weihenstephan

DNA in ng/mg FM



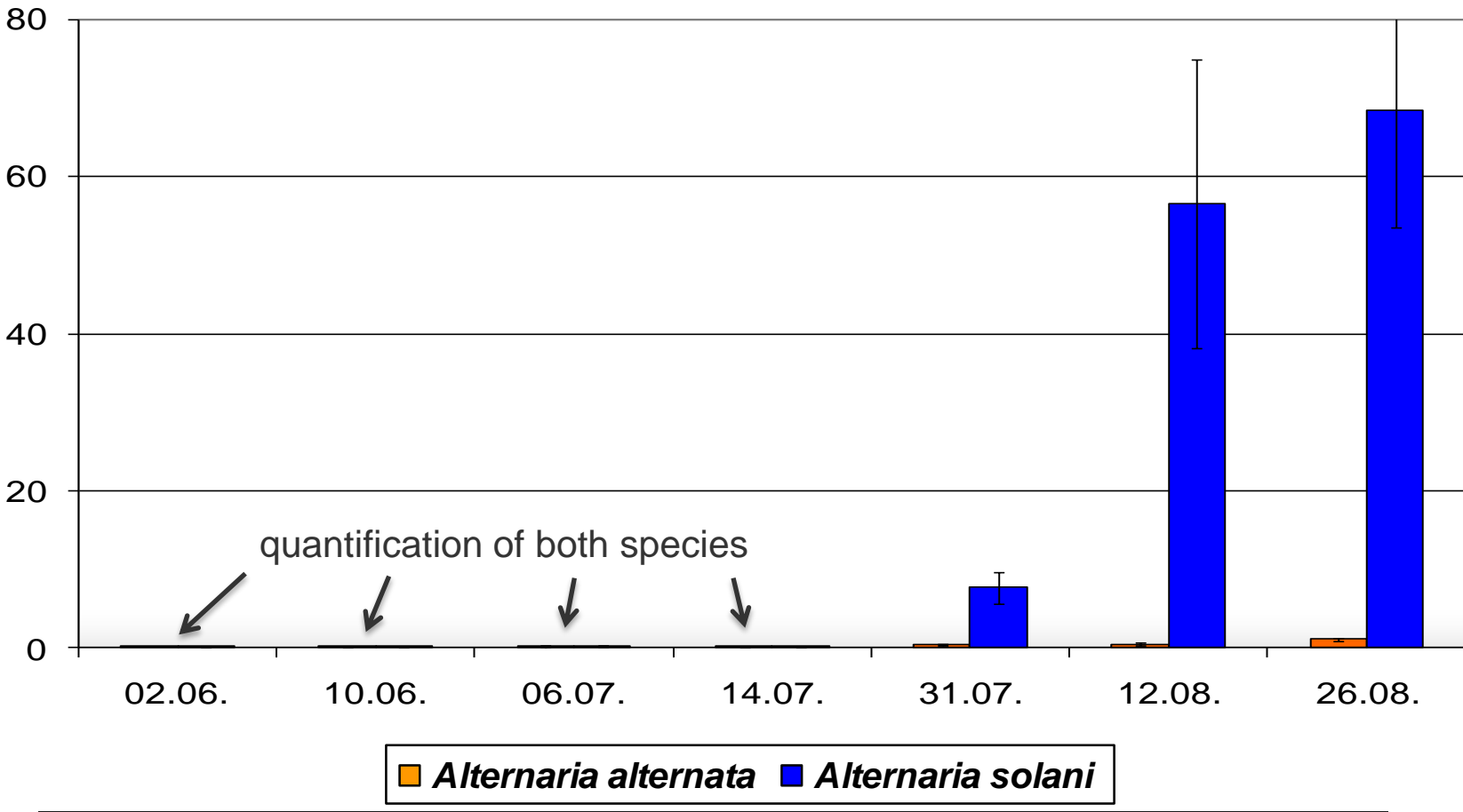
2004, Weihenstephan

DNA in ng/mg FM



2005, Weihenstephan

DNA in ng/mg FM



- Heavy early blight infections and epidemics throughout the past years
- Better insight into pathogen distribution through quantitative analysis (qPCR)
- Years with either more *A. solani* or *A. alternata*
- Earlier occurrence of *A. alternata* in 2003, detection of *A. solani* not before August
- Equal infestation by *A. solani* and *A. alternata* in 2004
- more severe and earlier appearance of *A. solani* in 2005



The differential success of a population or species over time is proportional to its genetic variation = genetic diversity



- better insight into early blight distribution and population structure
- investigation of *A. solani* population in the field
- better understanding of disease progression and improvement of disease control

Random Amplified Polymorphic DNA (RAPD) analysis as a tool to study genetic diversity

- the RAPD technique uses a single arbitrary 10mer as a primer for PCR to amplify genomic DNA fragments. No sequence information is required
- The primer anneals to the template DNA at several positions, which generates a specific DNA profile that can be used for genotype differentiation
- different individuals gives different PCR products,

primer: OPD-1: 5-ACC GCG AAC C-3



5-ACC GCG AAC C-3

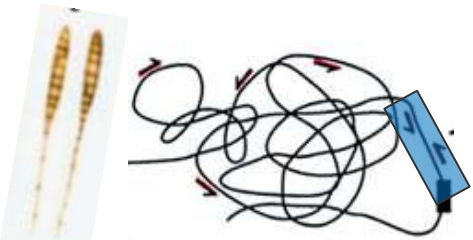


DNA A 3- TAC CTA TAA **TGG CGC TTG** GTA AGC GAT-5

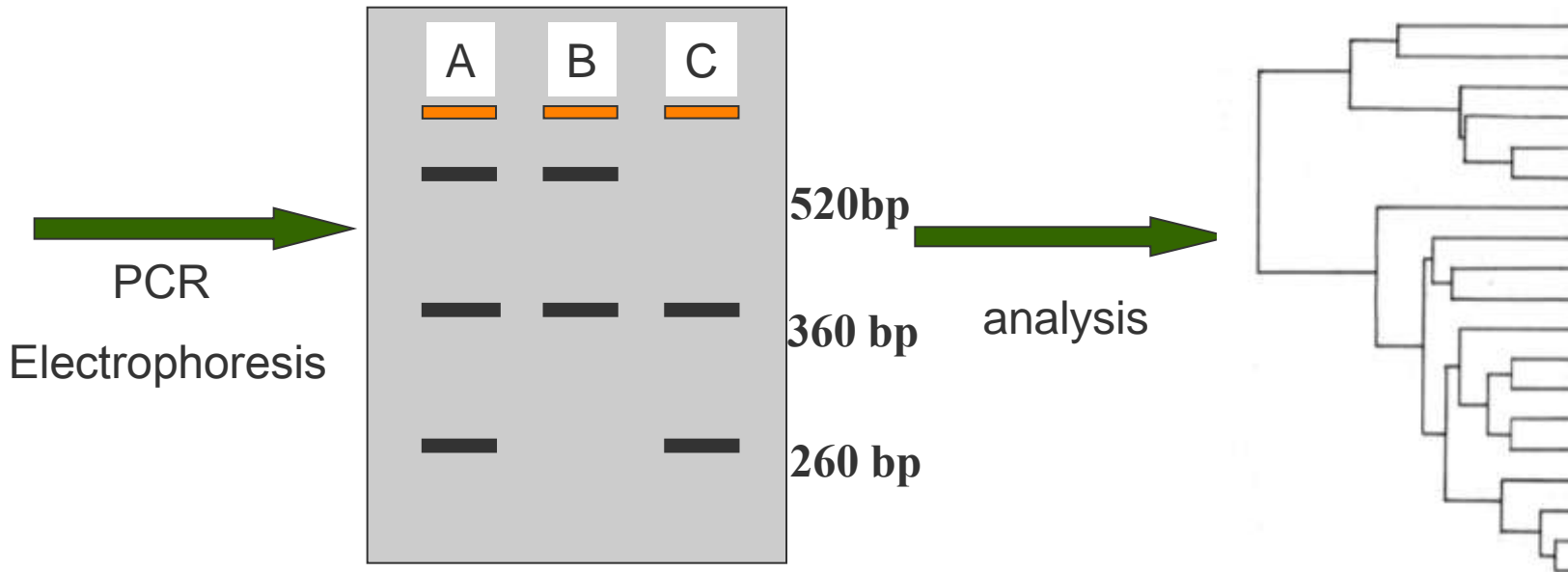
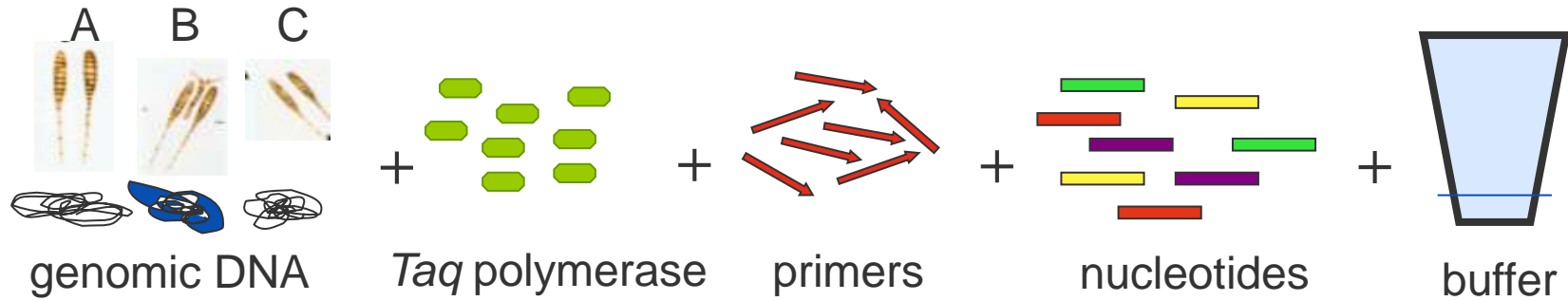
5-ACC **GCG** AAC C-3



DNA B 3- TAC CTA TAA **TGG TAC** TTG GTA AGC GAT-5



Random Amplified Polymorphic DNA (RAPD)

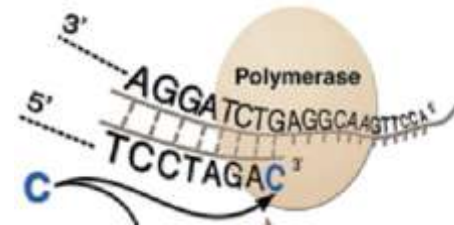


- isolation of 20 single spore isolates of *A. solani* (Weihenstephan, 2006, 2008)
- eight further isolates of *A. solani* from different German locations in 2008
- analysis with random primers (OP Operon Technologies, California)
~ test of 5 complete sets of OP random primers
- app. 120 different primers (OPA1-20, OPB 1-20, OPC-F...)
- 10 primers proved useful for differentiation of *Alternaria* genotypes

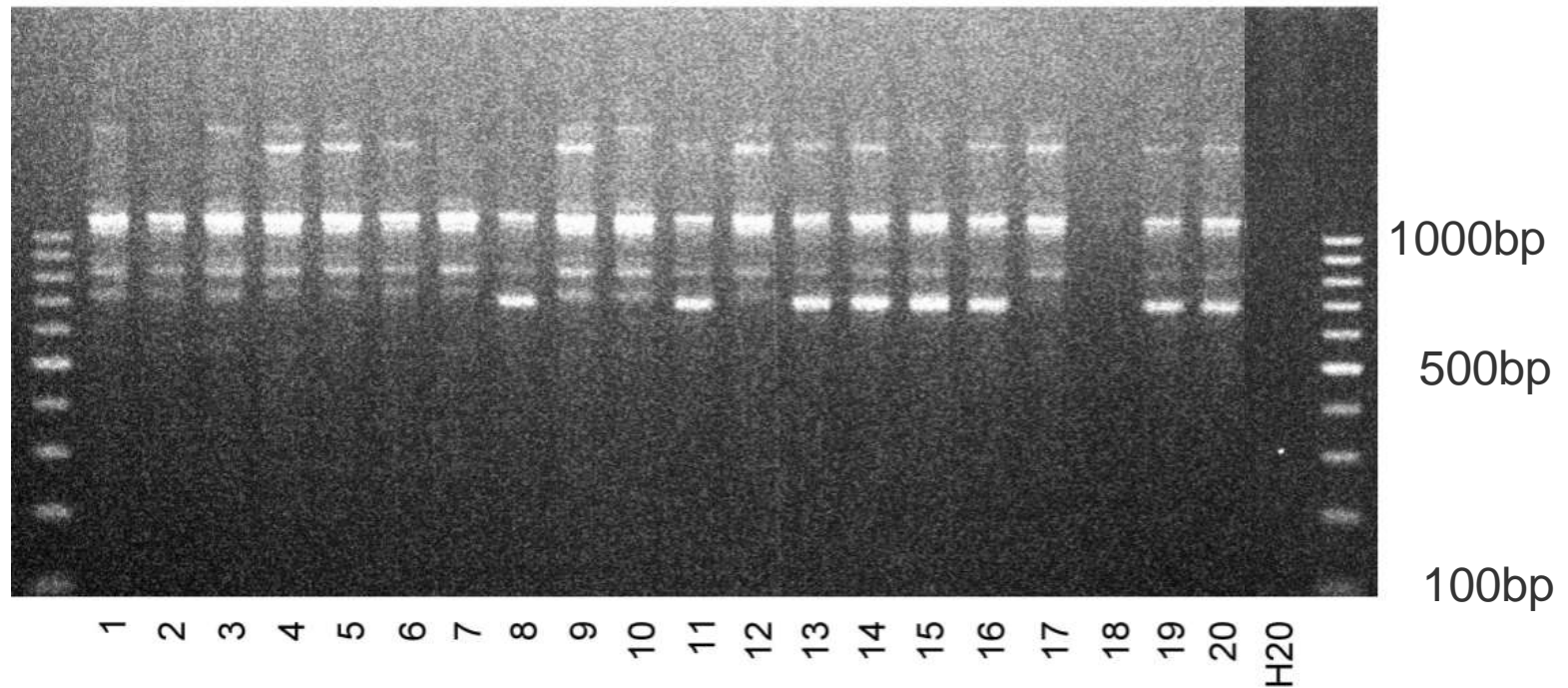
OPC 5, OPC 6, OPC 9, OPC 10, OPC 11, OPC 14, OPC 19,

OPD1, OPD2, OPD 11

- similarity matrix



Weihenstephan, 2008, comparison of 20 isolates of *A. solani* with OPD-1



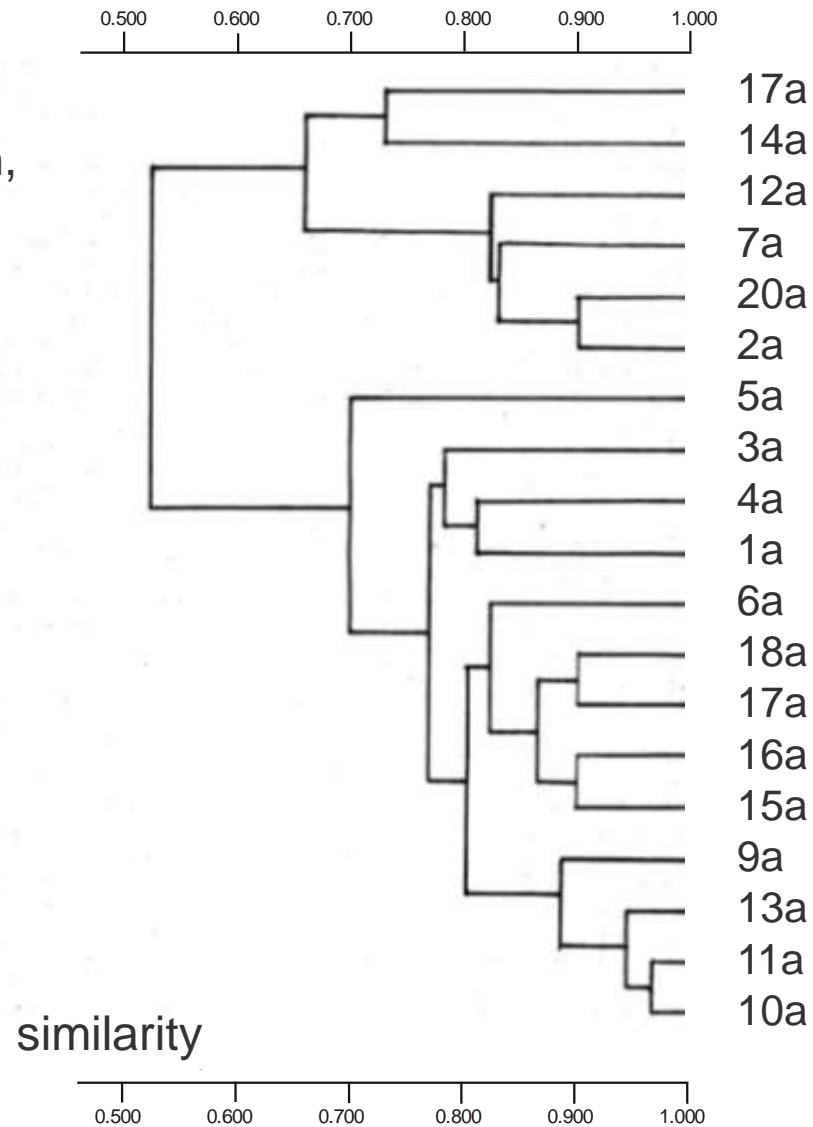
DNA of *Alternaria solani* isolates amplified with RAPD-primer OPD-1.

OPD-1: 5-ACCGCGAACC-3

Different band patterns are visible.

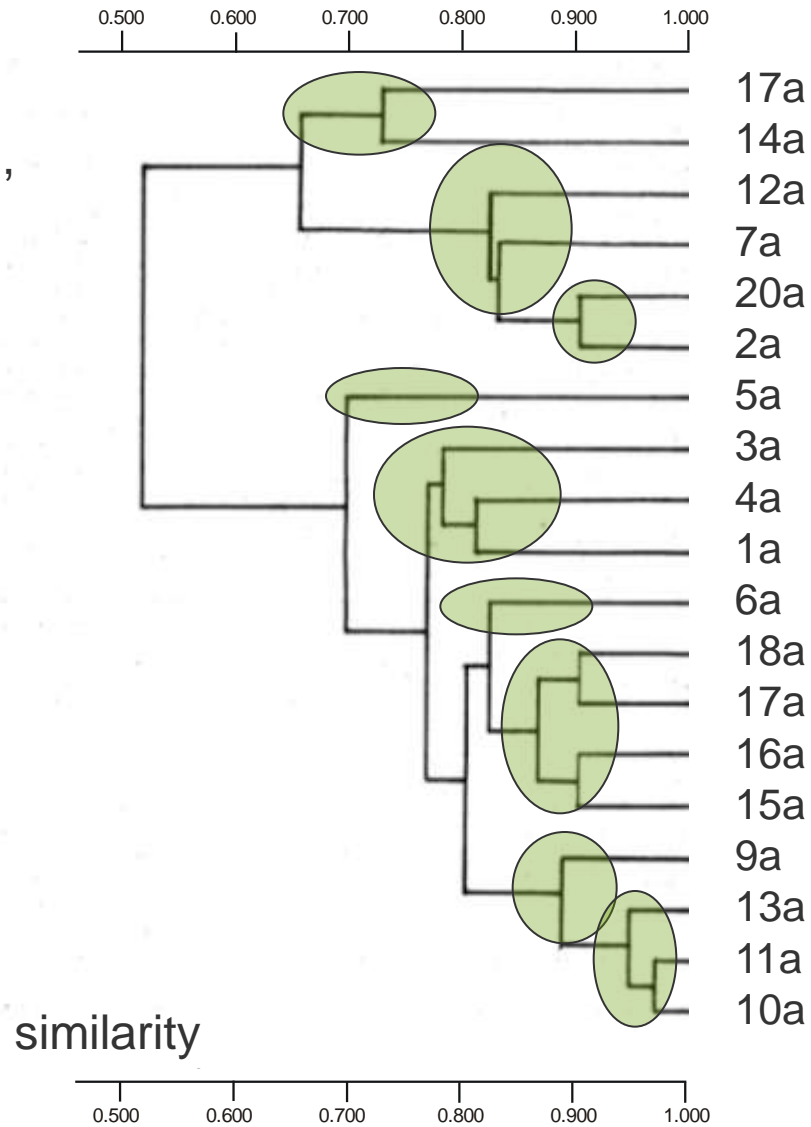
Results RAPD 2006

Weihenstephan,
2006

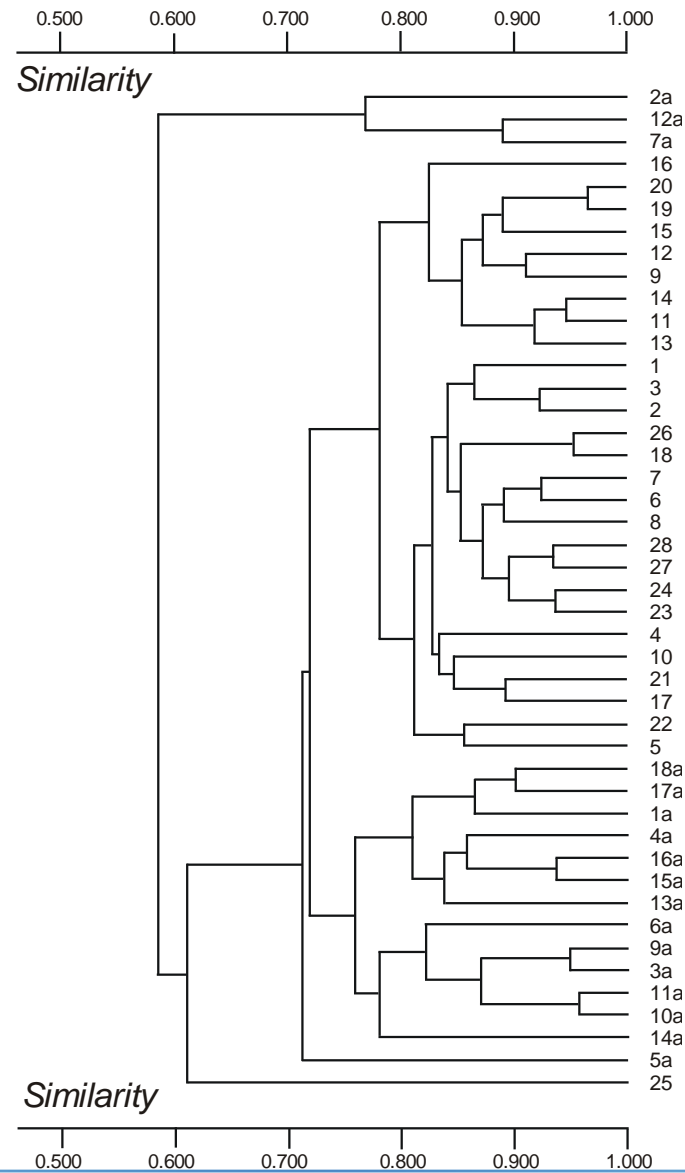


Results RAPD 2006

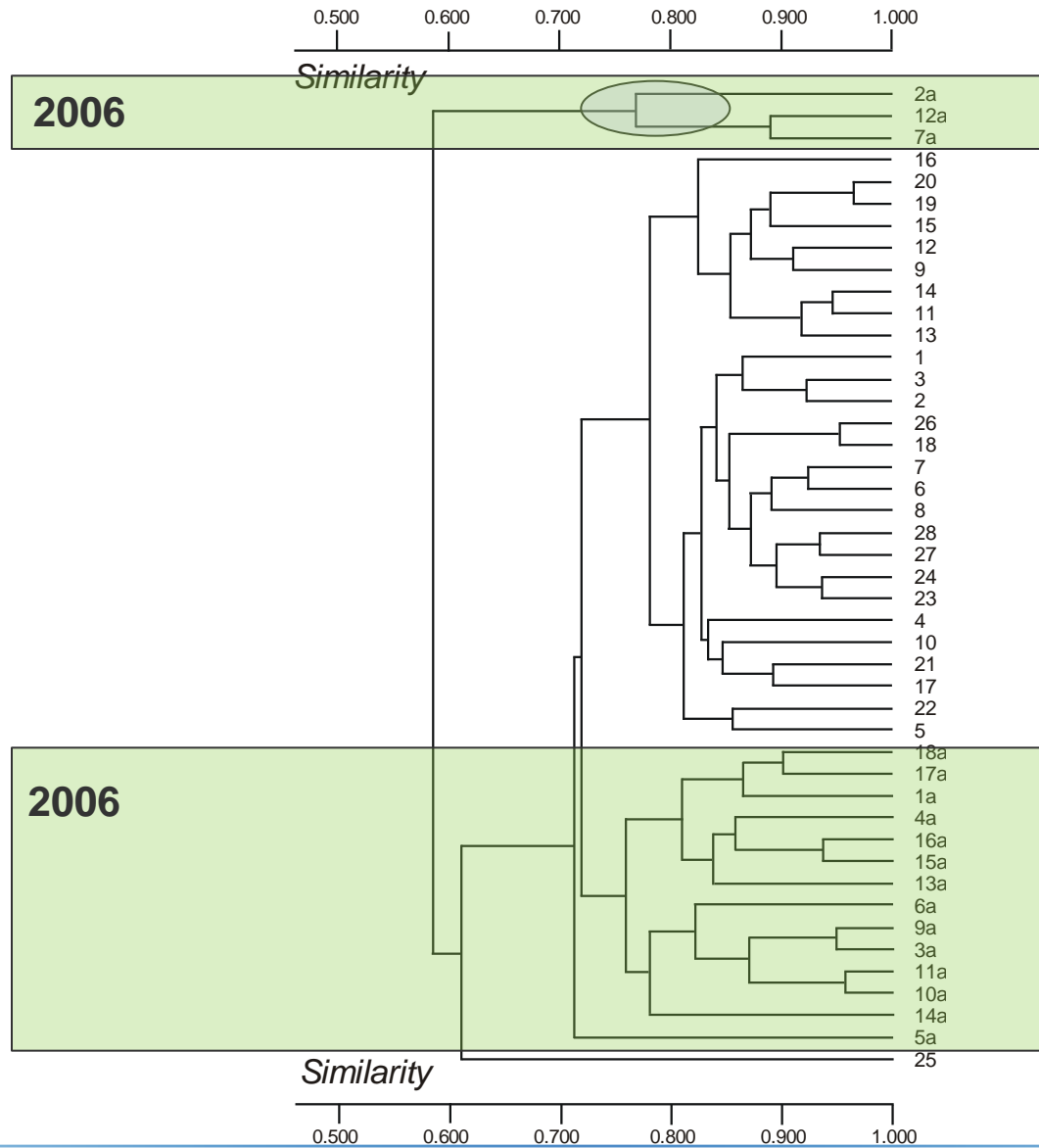
Weihenstephan,
2006



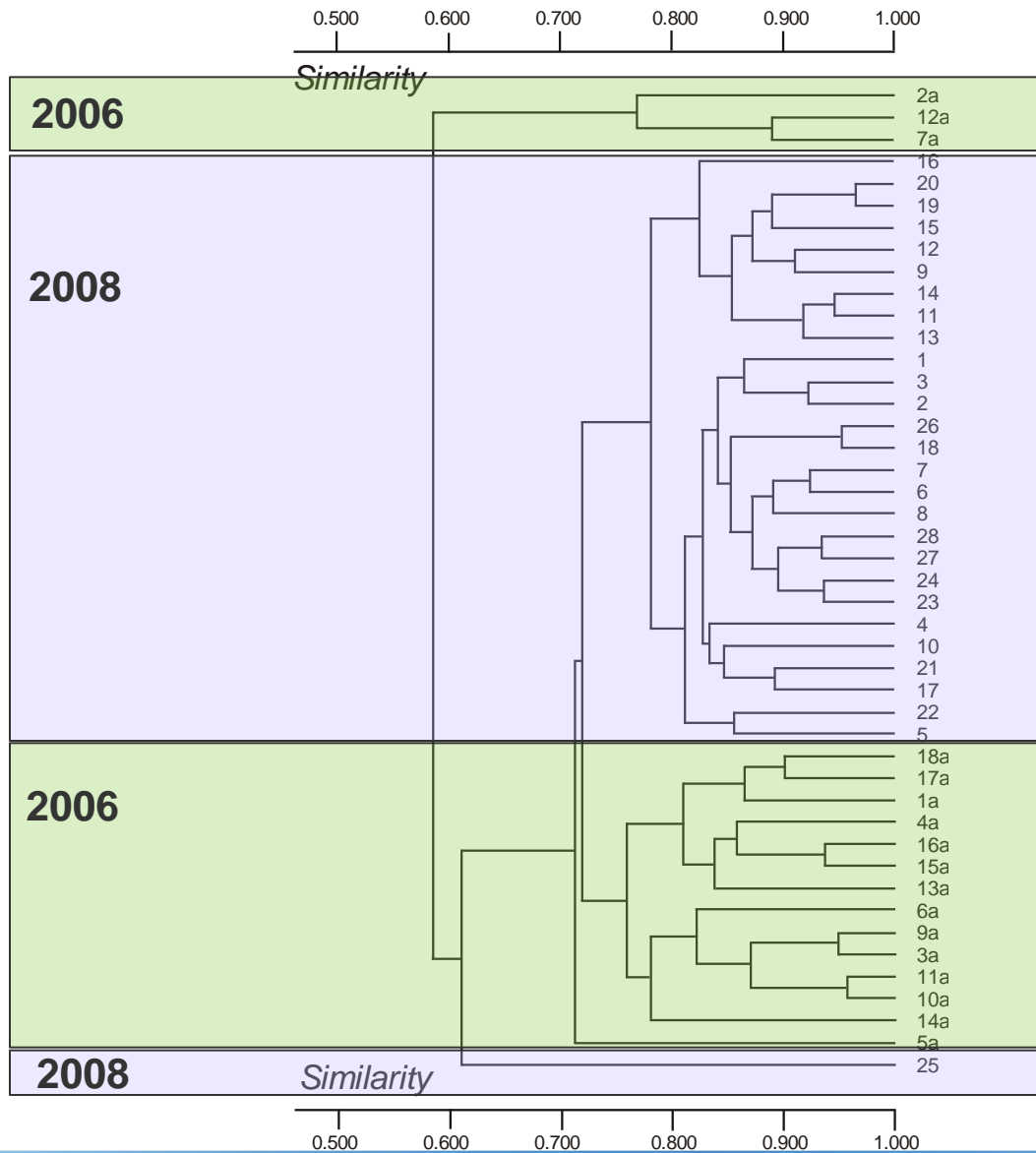
Results RAPD 2006 + 2008



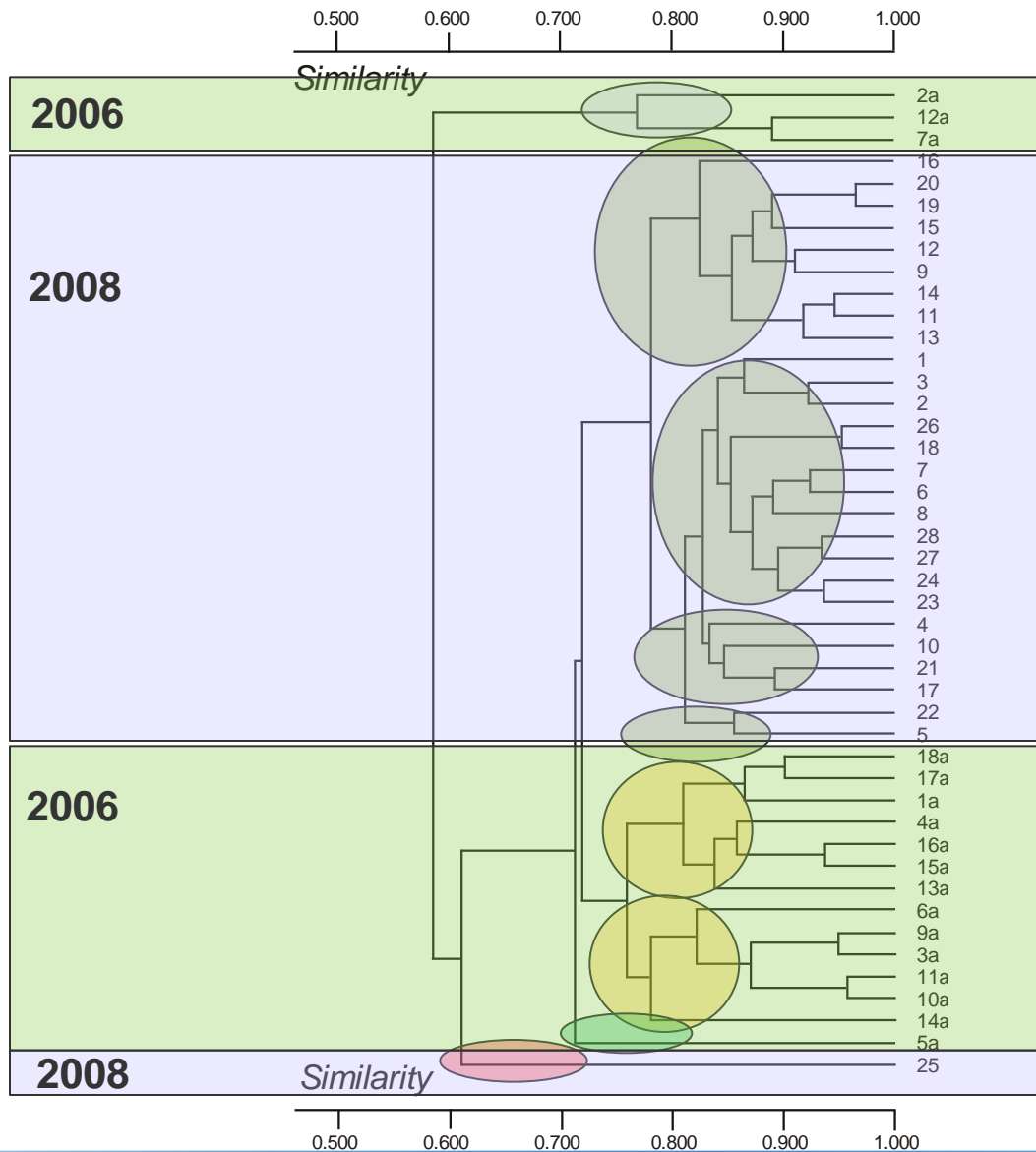
Results RAPD 2006 + 2008



Results RAPD 2006 + 2008



Results RAPD 2006 + 2008

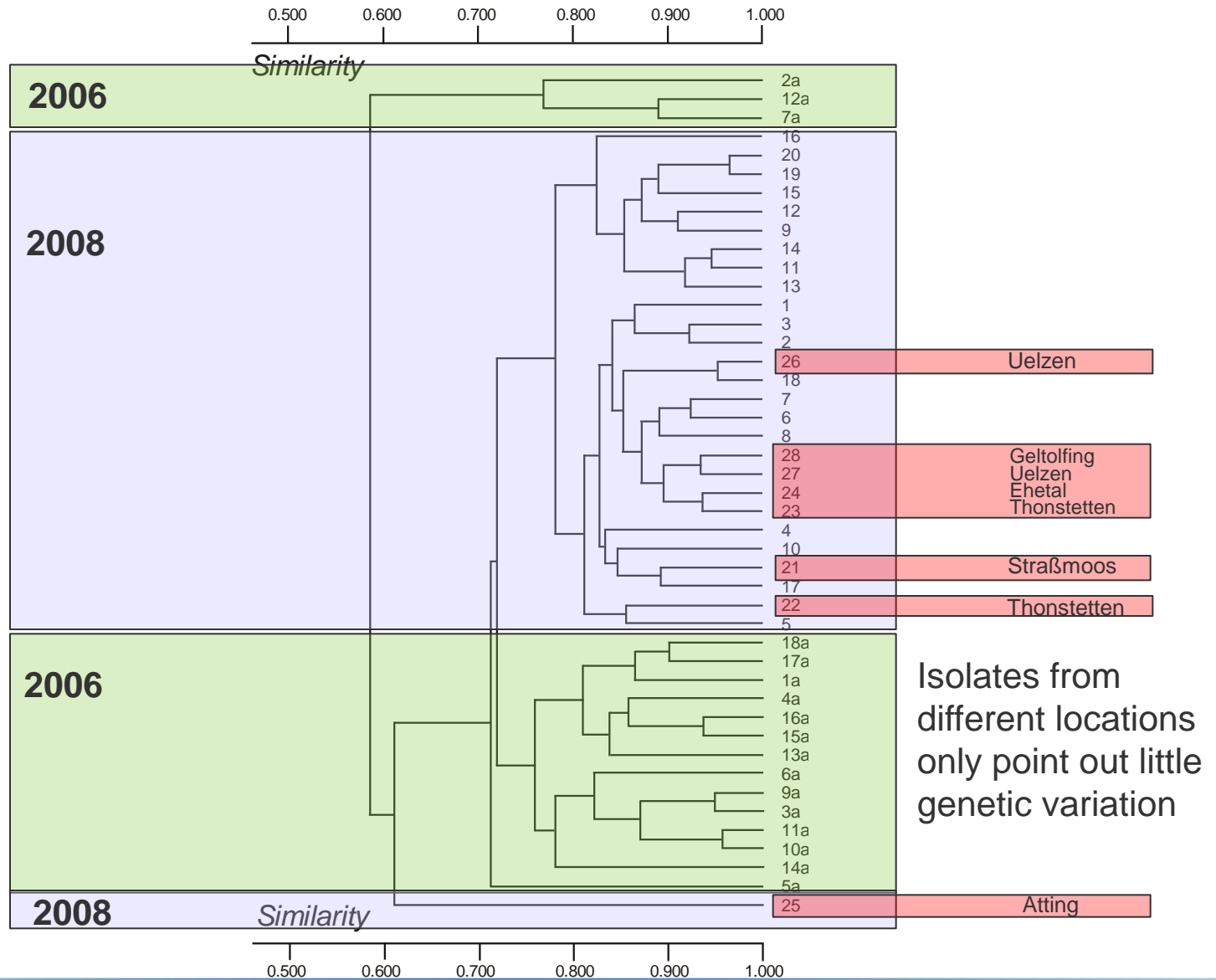


Completely different clusters in 2006 and 2008

No clustering according to location



Results RAPD 2006 + 2008



- high levels of heterogeneity among isolates in different years
- significant differences among isolates of one particular geographic origin
- Various geographical subpopulations are not genetically isolated
- *A. solani* is a highly variable pathogen
- Diversity due to natural mutation or large population size (McDonald & Linde, 2002)
- Mutation is the driving force facilitating strain evolution, virulence factor variation (Lourenco et al., 2009)
- Causal agent of early blight disease is able to adapt to changing environments or agricultural practices which can vary from year to year

“Wild species must have available a pool of genetic diversity if they are to survive environmental pressures If this is not the case, extinction would appear inevitable.”

(Otto Frankel 1983)



- high ability to adapt to changing environment facilitates build up of new populations, even within a small area
- preventive application of fungicide at an early disease stage (first application 7-8 weeks after crop emerge)
- monitor fields – note disease incidence & severity
- initial spray applications should not be carried out at disease progression (10-15% disease severity)
- prevent early blight by the use of few applications
- use full dose rates
- high genetic diversity supports selection of more virulent or fungicide resistant genotypes
- present potent fungicides (Strobilurines) with high resistance risk
- loss of sensitivity against QoI Inhibitors (F129L) Strobilurine within few years (USA)
- use combinations of single-site & multi-site fungicides

Thank you for your attention!

