

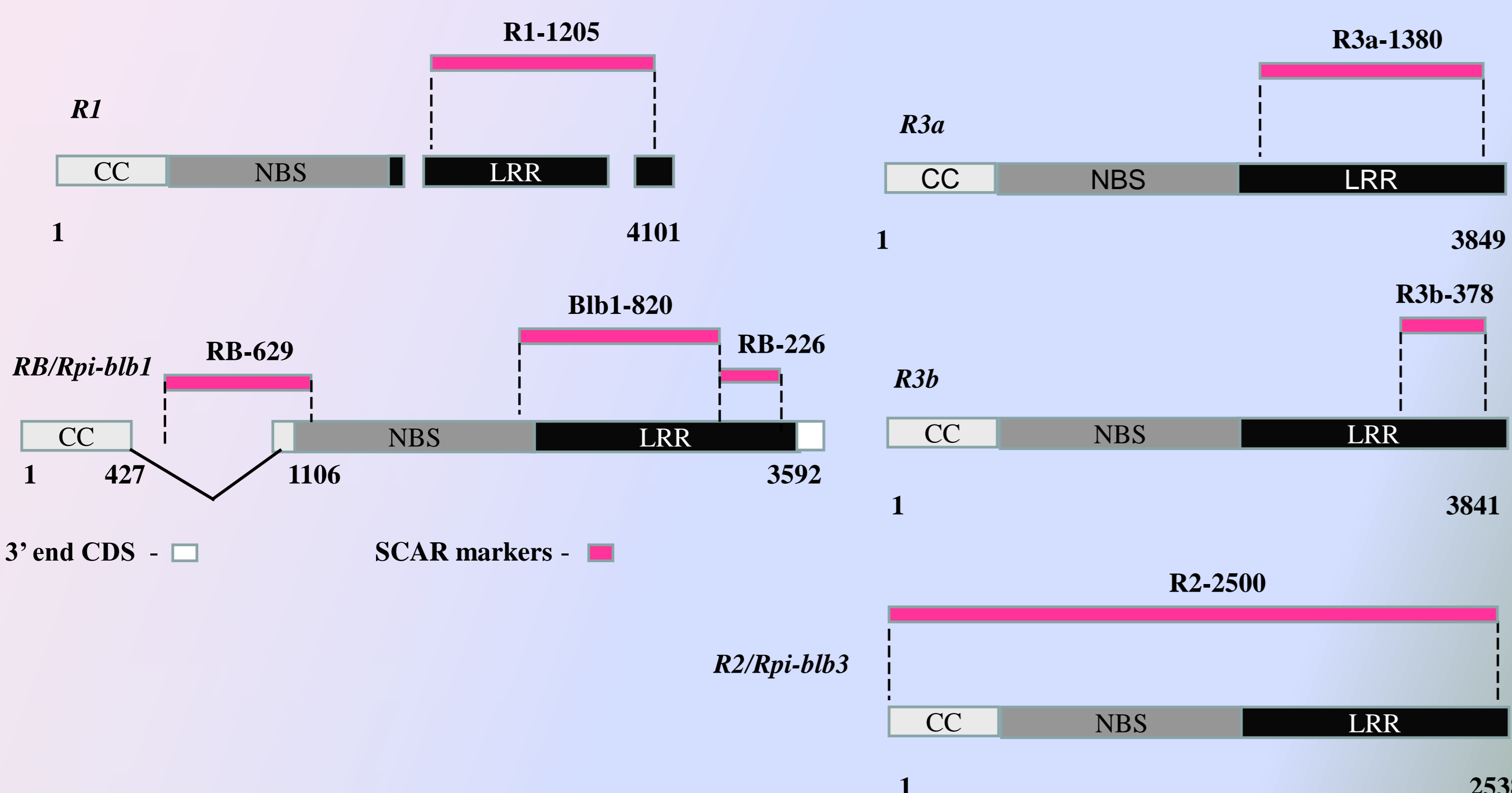
Structural homologues of CC-NBS-LRR genes for potato late blight resistance in wild *Solanum* species

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New races of *Phytophthora infestans* rapidly defeat potato late blight (LB) resistance based on germplasm transferred from *Solanum demissum*, so breeders search for new sources of durable LB resistance in genetic collections of wild *Solanum* species. SCAR markers for five race-specific genes initially characterized in *S. demissum* and *S. bulbocastanum*: *R1*, *R2/Rpi-blb3*, *R3a*, *R3b* and *RB/Rpi-blb1* – were employed to screen a collection of wild *Solanum* accessions representing six series from section Petota and clones developed from these accessions. The pattern thus produced will facilitate further mining for new *R* genes for LB resistance and promote the evolutionary studies of CC-NBS-LRR resistance genes.

Markers and their sizes, bp	Prototype clone	Chromosome	Position in the prototype clone	References
RB-629	AY336128	8	595-1223	[1]
RB-226	AY336128	8	3143-3368	[2]
Blb1-820	AY336128	8	2547-3143	[3]
R1-1205	AF447489	5	5126-6331	[1]
R2-2500	FJ536325	4	1-2538	[4]
R3a-1380	AY849382	11	1677-3056	[1]
R3b-378	JF900492	11	94818-95195	[4]



SCAR markers for *R* genes for LB resistance. Primer positions (base pairs) are shown against the structures of CC-NBS-LRR kinases for prototype genes *R1*, *R2/Rpi-blb3*, *R3a*, *R3b* and *RB/Rpi-blb1*.

When using this evidence for selecting the genotypes of paramount interest for further breeding program, the crucial problem is the functional identity of newly found CC-NBS-LRR homologues. In some cases, we have supported the evidence for new alleles of the *R* genes in orphan *Solanum* species by cloning and sequencing [1], while other laboratories provide even more convincing data from co-expression studies in *Nicotiana benthamiana* and potato transformation.

Genes-prototype	SCAR markers found in species	Mapped and cloned genes and their homologues	Genes cloned from the following species	Co-expression or transient complementation in <i>N. benthamiana</i>	Transformation of potato
<i>R1</i>	<i>dms</i> , <i>hou</i> , <i>plt</i> , <i>sto</i> , <i>ber</i> , <i>mcd</i>	<i>R1</i> [11,12]	<i>dms</i> [11,12]	-	<i>R1 dms</i> [11]
<i>R2/Rpi-blb3</i>	<i>blb</i> , <i>hou</i> , <i>pnt</i>	<i>R2/Rpi-blb3</i> <i>Rpi-abpt</i> <i>R2-like</i> [8] <i>Rpi-mcd1</i> [9] <i>Rpi-edn1.1</i> <i>Rpi-hjt1.1</i> , <i>Rpi-hjt1.2</i> , <i>Rpi-hjt1.3</i> <i>Rpi-snk1.1</i> , <i>Rpi-snk1.2</i> [10]	<i>tub</i> , <i>dms</i> , <i>edn</i> , <i>snk</i> <i>hjt</i> , <i>blb</i> , <i>vrn</i> , <i>mcd</i> [8-10]	<i>R2 Rpi-blb3</i> <i>Rpi-abpt</i> <i>R2-like</i> [8] <i>Rpi-mcd1</i> [9]	<i>R2 Rpi-abpt</i> <i>R2-like</i> [8] <i>Rpi-mcd1</i> [9]
<i>R3a</i>	<i>blb</i> , <i>hou</i> , <i>dms</i> , <i>cph</i> , <i>pld</i> , <i>mcd</i> , <i>plt</i> , <i>sto</i>	<i>R3a</i> [13] <i>Rpi-sto2</i> [10] <i>R8</i> [19] <i>R3b</i> [14] <i>R5</i> , <i>R6</i> , <i>R7</i> , <i>R9</i> , <i>R10</i> , <i>R11</i> [13]	<i>tub</i> , <i>dms</i> [6], <i>sto</i> [10,1], <i>blb</i> , <i>cph</i> , <i>hou</i> , <i>plt</i> [1]	<i>Rpi-sto2</i> , <i>dms</i> , <i>sem</i> , <i>mcd</i> , <i>sto</i> , <i>cph</i> , <i>ehr</i> [10]	
<i>R3b</i>	<i>blb</i> , <i>hou</i> , <i>dms</i> , <i>cph</i> , <i>hjt</i> , <i>jam</i> , <i>pnt</i> , <i>ver</i> , <i>sto</i> , <i>plt</i>	<i>R3b</i> [14] <i>R3a</i> [13] <i>R8</i> [19] <i>R5</i> , <i>R6</i> , <i>R7</i> , <i>R9</i> , <i>R10</i> , <i>R11</i> [13]	<i>dms</i> [8]	-	
<i>RB/Rpi-blb1</i>	<i>blb</i> , <i>hou</i> , <i>dms</i> , <i>cph</i> , <i>hjt</i> , <i>jam</i> , <i>pnt</i> , <i>ver</i> , <i>pld</i> , <i>ber</i> , <i>sto</i> , <i>plt</i> , <i>mcd</i>	<i>RB/Rpi-blb1</i> [15,16] <i>Rpi-bt1</i> [17] <i>RB ver</i> [6] <i>Rpi-sto1</i> , <i>Rpi-pta1</i> , <i>Rpi-plt1</i> [3]	<i>blb</i> , <i>sto</i> , <i>ver</i> , <i>plt</i> [3,6,15-18]		<i>RGA3</i> [15], <i>RB</i> [16, 18], <i>Rpi-bt1</i> [17], <i>RBver</i> [6]

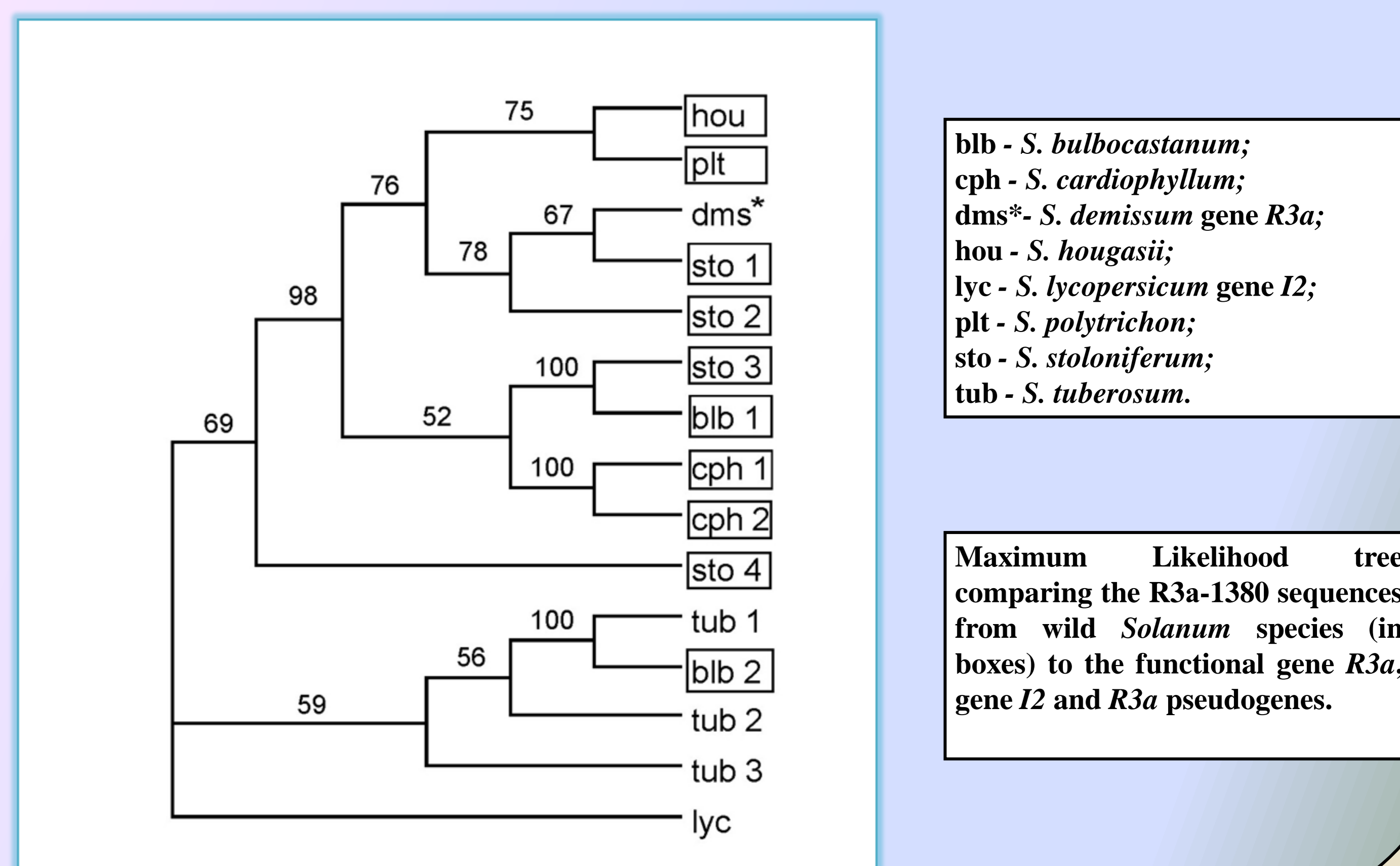
ber - *S. berthaultii*, *cph* (*ehr*) - *S. cardiophyllum*=*S. ehrenbergii*, *dms* - *S. demissum*, *edn* - *S. edinense*, *hou* - *S. hougasii*, *hjt* - *S. hjertingii*, *jam* - *S. jamesii*, *tbr* - *S. tuberosum*, *mcd* - *S. microdontum*, *pnt* - *S. pinnatisectum*, *plt* - *S. polytrichon* = *S. stoloniferum*, *pld* - *S. polyadenium*, *sem* - *S. semidemissum*, *snk* - *S. schenckii*, *sto* - *S. stoloniferum*, *tbr* - *S. tuberosum*, *vrn* - *S. vernei*, *ver* - *S. verrucosum*.

SCAR markers for CC-NBS-LRR resistance genes have been employed to screen 218 *Solanum* accessions of 14 wild *Solanum* species. Structural homologues of particular *R* genes widely differ in their distribution patterns in the section Petota. The genes *R1*, *R2*, *R3a* and *R3b* initially identified in *S. demissum* were found in many taxonomically distant species. Structural homologues of the *RB/Rpi-blb1* gene, which was initially identified in genome B of *S. bulbocastanum* and *S. stoloniferum* (for review see [5]), were found in many genome A species, and only *RBver* in *S. verrucosum* was found to participate in LB resistance [6]. Such evidence suggests that many *R*-gene structures evolved before the divergence of genomes A and B and subsequent *Solanum* speciation. The presence of markers RB-226 and RB-820 consistently corresponds to the presence of the functional gene *Rpi-blb1*, whereas marker RB-629 presents its structural homologue poorly related to LB resistance (for further details see the poster by Fadina et al. at this meeting).

Series and species [20]	The total number of screened accessions	Occurrence of <i>R</i> -gene markers						
		R1-1205	R2-2500	R3a-1380	R3b-378	RB-629	RB-226	Blb1-820
<i>Bulbocastana</i> (<i>S. bulbocastanum</i>)	28	0						
<i>Demissa</i>	<i>S. demissum</i>	38	nd					0
	<i>S. hougasii</i>	7						0
<i>Longipedicellata</i>	<i>S. hjertingii</i>	6	0	0			0	nd
	<i>S. polytrichon</i>	12		0			nd	nd
	<i>S. stoloniferum</i>	51		nd				
<i>Pinnatisecta/ Cardiophylla</i>	<i>S. cardiophyllum</i>	7	0	nd			0	nd
	<i>S. ehrenbergii</i>	17	0				0	0
	<i>S. jamesii</i>	13	0	0	0		0	nd
	<i>S. pinnatisectum</i>	10	0		0		0	0
<i>Polyadenia</i>	<i>S. polyadenium</i>	8	0	nd	0		0	nd
	<i>S. verrucosum</i>	12	0	0	0	0	0	0
<i>Tuberosa</i>	<i>S. berthaultii</i>	7		nd	0		0	nd
	<i>S. microdontum</i>	6		0	0		0	nd

■ - frequent occurrence of the marker □ - less frequent occurrence of the marker

Cloned markers R1-1205 in *S. polytrichon* and *S. stoloniferum* and R3a-1380 in *S. bulbocastanum*, *S. cardiophyllum*, *S. hougasii*, *S. polytrichon* and *S. stoloniferum* were 98-99% identical to the corresponding regions in the prototype genes. The phylogenetic analysis of the R3a-1380 fragments from wild *Solanum* species sequenced in our laboratory when compared to those of the *demissum* *R3a* gene, tomato *I2* gene and *R3a*-like inactive homologues from the NCBI Genbank demonstrated that the R3a-1380 sequences belonged to the same cluster as *R3a* presuming that they represented the active genes [1].

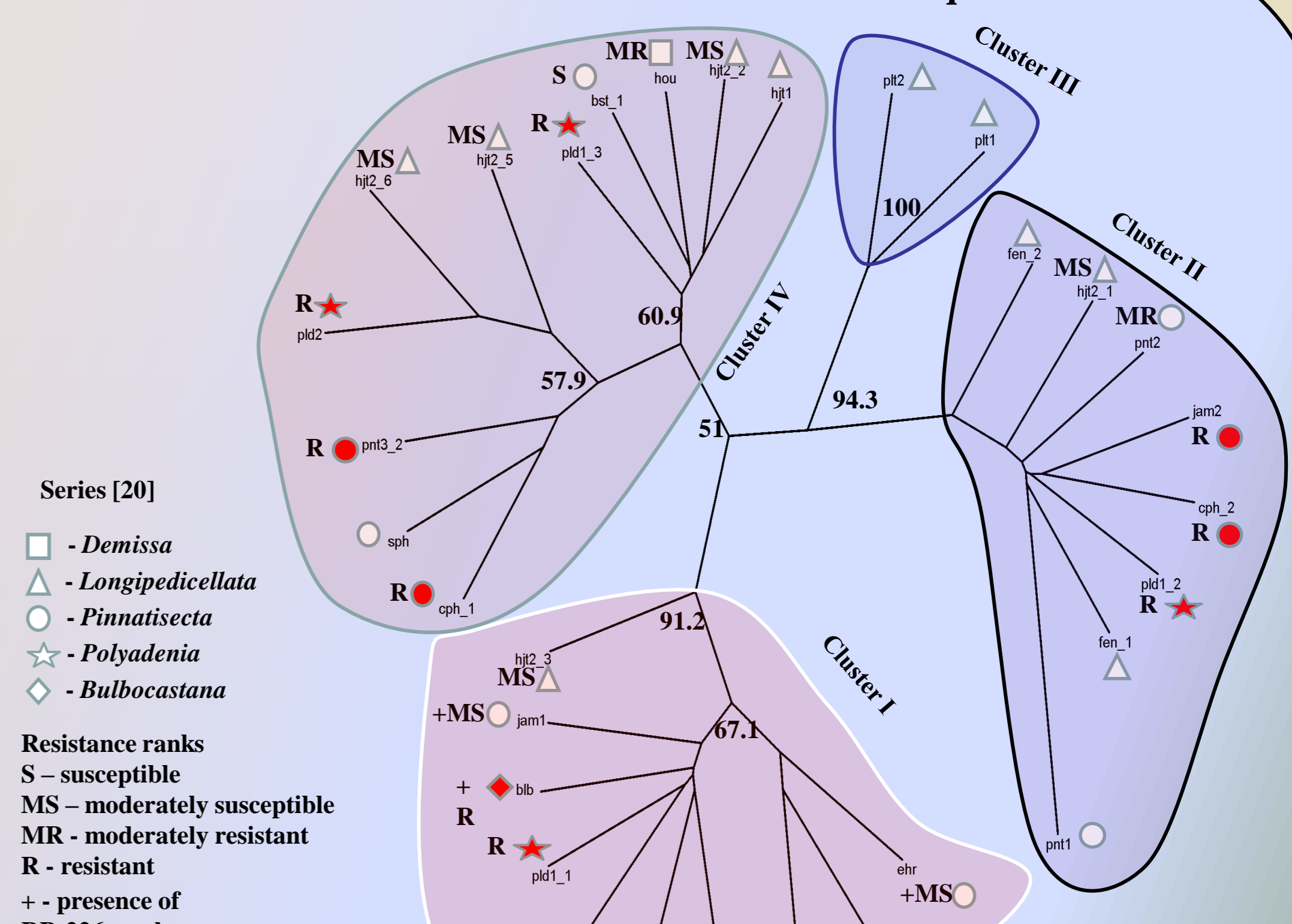


blb - *S. bulbocastanum*;
cph - *S. cardiophyllum*;
dms* - *S. demissum* gene *R3a*;
hou - *S. hougasii*;
lyc - *S. lycopersicum* gene *I2*;
plt - *S. polytrichon*;
sto - *S. stoloniferum*;
tub - *S. tuberosum*.

Maximum Likelihood tree comparing the R3a-1380 sequences from wild *Solanum* species (in boxes) to the functional gene *R3a*, gene *I2* and *R3a* pseudogenes.

When RB-629 was cloned from 16 accessions representing 12 wild *Solanum* species, the pattern of polymorphisms was neither species- nor series-specific, and we conclude that observed diversity of *RGA2*-like loci emerged before *Solanum* speciation. The marker RB-226 was found only in the cluster I genotypes. Marker presence and particular haplotypes were not immediately associated with high LB resistance [7].

Maximum-likelihood tree of *RB*-like sequences



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